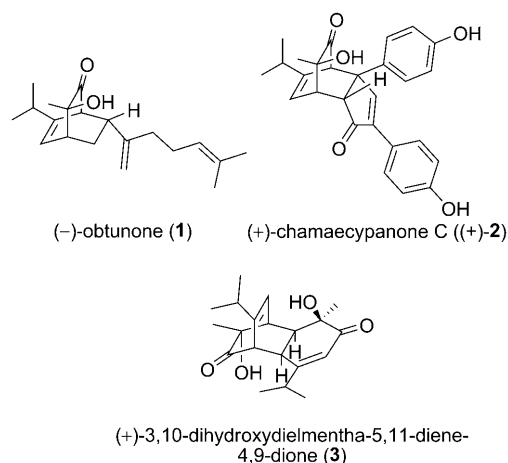


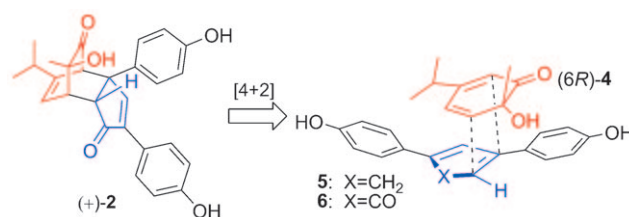
Enantioselective Synthesis of (+)-Chamaecyanone C: A Novel Microtubule Inhibitor**

Suwei Dong, Ernest Hamel, Ruoli Bai, David G. Covell, John A. Beutler, and John A. Porco, Jr.*

A number of bicyclo[2.2.2]octenone-containing natural products have been isolated from the heartwood of *Chamaecyparis obtusa* var. *formosana* (Scheme 1), including the Diels–Alder (DA) adducts^[1] obtunone (**1**)^[2] and chamaecyanone C (**2**)^[3] as well as the [4+2] dimer (+)-**3**.^[2,4] (+)-**2** has been shown to exhibit potent cytotoxicity against several human cancer cells, including human oral epidermoid carcinoma (KB; IC₅₀ = 190 nM).^[3] The biosynthesis of **2** is proposed^[3] to occur through *endo* [4+2] cycloaddition between cyclohexa-2,4-dienone **4** (Scheme 2) and 1,3-bisaryl cyclopenta-1,3-diene **5**, and subsequent oxidation to an enone—in accordance with literature reports of cyclopentadienes as biosynthetic precursors to natural products.^[1b] An alternative biosynthetic possibility involves the corresponding cyclopentadienone **6** as the dienophile, and which may also be considered in light of known biosyntheses that involve reactive cyclopentadienones.^[5] Herein, we report a concise synthesis of both enantiomers of chamaecyanone C (**2**). The synthesis involves a retro-Diels–Alder/Diels–Alder cascade of dimer **3**, which is obtained by utilizing copper-mediated asymmetric oxidative dearomatization.^[6] Also presented are biological studies which document that the cytotoxic action of (+)-chamaecyanone C involves mitotic arrest as a consequence of its binding in the colchicine site of tubulin.



Scheme 1. Representative bicyclo[2.2.2]octenone-containing natural products.



Scheme 2. Plausible biosyntheses of (+)-chamaecyanone C ((+)-**2**).

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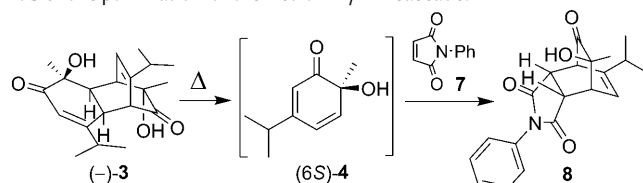
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Inspired by literature reports on tandem retro-DA/DA reactions of dimers derived from *ortho*-quinols and masked *ortho*-benzoquinones (MOBs),^[7,8] we first evaluated reactions between the readily accessible dimer (–)-**3**^[6] and *N*-phenylmaleimide (**7**) under thermolytic reaction conditions in different solvents (Table 1). Although reactions in toluene (Table 1, entry 1) and chlorobenzene (Table 1, entry 2) generated the desired cycloadduct **8** in moderate to good conversion (12 h), reactions in mesitylene at 150°C gave both excellent conversion and yield of isolated **8** in 1.5 hours (Table 1, entries 3 and 4).

By using these optimized reaction conditions, a number of representative dienophiles were thermolyzed in the presence of dimer (–)-**3** in mesitylene (Table 2). Thermolysis reactions with methyl vinyl ketone (**9**; Table 2, entry 1), 2,3-dihydrofuran (**10**; Table 2, entry 2), and indene (**11**; Table 2, entry 3) successfully generated bicyclo[2.2.2]octenones **12–14** in good to excellent yields. These results underscore the reactivity of *ortho*-quinols as both “normal” and “inverse-demand”

Table 1: Optimization of the Retro-DA/DA Cascade.


Entry	Solvent	T [°C]	Dienophile 7 [equiv]	t [h]	Conv. [%] ^[a]
1	toluene	110	5	12	69
2	chlorobenzene	130	5	12	92
3	mesitylene	150	5	1.5	> 99 (98) ^[b]
4	mesitylene	150	3	1.5	> 99 (97) ^[b]

[a] Conversion is based on ¹H NMR analysis of **8** and starting material (–)-**3**. [b] Yield of isolated **8**.

Table 2: Tandem retro-DA/DA reactions using bicyclooctenone (–)-**3**.^[a]

Entry	Dienophile	Equiv	Cycloadduct	t [h]	Yield [%] ^[b]
1		5		3	92
2		20		12	84
3		10		3	99
4 ^[d]		10		4	42
				4	39
5		5		4	98
6		2.5		5	57

[a] Reaction conditions: dimer (–)-**3**, dienophile, mesitylene, 150 °C. [b] Yield of isolated product after column chromatography. [c] Approximately 6% of an inseparable minor product was detected by ¹H NMR spectroscopy. [d] Acetylation was required for product separation.

dienes. The observed regioselectivity for products **12–14** is in agreement with those reported for related cyclohexadienones (MOBs).^[7c,d] Treatment of (–)-**3** with β-myrcene (**15**; Table 2, entry 4) smoothly generated an inseparable 1:1 mixture of *ent*-obtunone (**1**) and a decalin product, both of which were

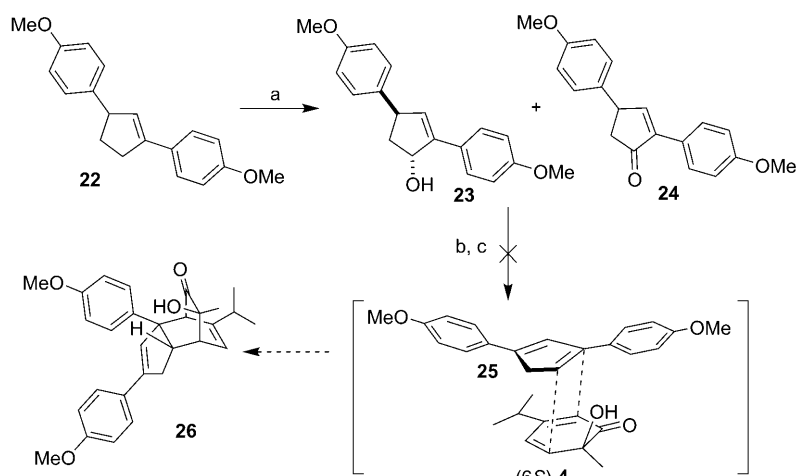
acetylated to afford **16** and **17**.^[9] Hydrolysis of **16** (aq. NaOH/MeOH) afforded optically pure *ent*-**1**.^[2,10] Furthermore, cyclopentadiene dimer **18** (Table 2, entry 5) was very reactive, and afforded the [4+2] adduct **19** as a single diastereomer in nearly quantitative yield.^[7a] In contrast, use of cyclopentadienone dimer **20** (Table 2, entry 6) produced **21** only in moderate yield, probably as a result of side reactions (including decarbonylation) of **20** at high temperature.^[11]

Based on our ability to trap (6*S*)-**4** with a number of dienophiles, we proceeded to evaluate both cyclopentadienes and cyclopentadienones for the synthesis of **2**. Accordingly, we targeted a single starting material for the preparation of both precursors. Starting from the known bisarylcyclopentene derivative **22**,^[12] allylic oxidation using selenium dioxide afforded alcohol **23** as the major product (50% yield) along with a small amount of enone **24** (Scheme 3). Although diaryl cyclopentadienes^[13] were detected by GC-MS analysis under acid-catalyzed dehydration conditions (cat. MP-TsOH, toluene, 110 °C, 1 h),^[14] all attempts to isolate pure product **25**, or trap it with reactive dienophiles (e.g. maleic anhydride, tetracyanoethylene) failed. Moreover, thermolysis of the crude mixture from either dehydration of **23** or base-promoted elimination of the derived mesylate derivative with dimer **3** also did not afford the desired cycloadduct **26**.

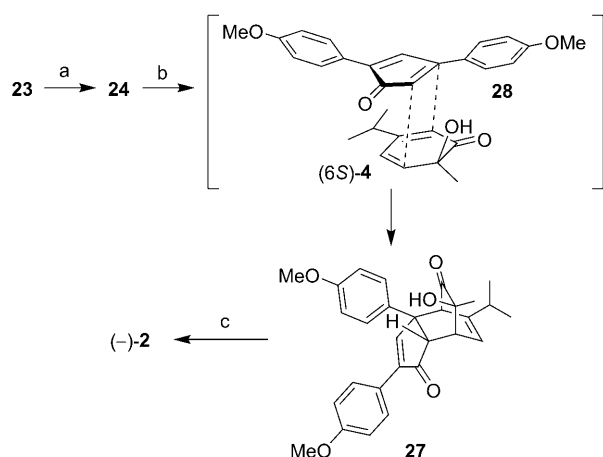
Alternatively, allylic alcohol **23** could be efficiently converted into cyclopentenone **24** using IBX as the oxidant^[15] (Scheme 4). After extensive experimentation, it was found that oxidation of **24** using DDQ^[16] in the presence of dimer **3** afforded the desired cycloadduct **27** in good yield. The *endo* configuration of **27** was unambiguously assigned by NOE experiments.^[10] The transformation presumably proceeds through the initial formation of the reactive cyclopentadienone **28** from cyclopentenone **24**.^[17] Unfortunately, all efforts to isolate either the cyclopentadienone monomer or derived dimers have thus far failed in control experiments. Finally, treatment of **27** with BBr₃ effected smooth demethylation to afford (–)-chamaecypa-*n*-one C ((–)-**2**; 86%). To the best of our knowledge, this is the first example of the generation of a 2,4-diarylcyclopentadienone and its usage in natural product synthesis.^[18] The instability and high reactivity of the diarylcyclopentadienone intermediate^[19] is likely due to the relief of antiaromaticity upon cycloaddition as suggested by Harmata et al.^[20]

In a similar manner, we prepared (+)-chamaecypa-*n*-one C ((+)-**2**; Scheme 5). Hydrogenation of **29** quantitatively generated 2,4-disubstituted phenol **30**. An asymmetric hydroxylation/α-ketol rearrangement/dimerization sequence^[6] afforded (+)-dimer **3** in moderate yield over two steps (> 99% *ee*), and which was further elaborated into (+)-chamaecypa-*n*-one C (53%, over 2 steps from enone **24**). Synthetic (+)-**2** was confirmed as being identical to natural chamaecypa-*n*-one C by comparison of ¹H and ¹³C NMR spectra, the mass spectrum, IR, and [α]_D data, thus confirming its absolute configuration.^[10]

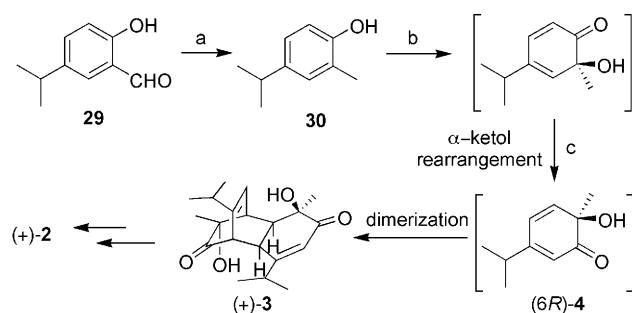
Both enantiomers of **2** were tested in the National Cancer Institute's 60-cell single dose assay at 10^{–5} M. Confirming earlier studies on the natural product,^[3] (+)-**2** inhibited tumor cell growth by an average of 71%, while (–)-**2** had no effect. (+)-**2** was then tested in a dose response format, where it



Scheme 3. Reagents and conditions: a) SeO_2 (0.5 equiv), TBHP (2 equiv), DCE, 60°C , 2 h, 50% (**23**), and 5% (**24**); b) cat. MP-TsOH, toluene, 110°C , 1 h; or Martin sulfuran, CH_2Cl_2 , RT, 0.5 h; c) $(-)\text{-3}$ (0.2 equiv), mesitylene, 150°C . DCE = 1,2-dichloroethane, MP = macroporous polymer, TBHP = *tert*-butyl hydroperoxide, Ts = 4-toluenesulfonyl.



Scheme 4. Reagents and conditions: a) IBX (2.0 equiv), toluene/DMSO (1 M, 2:1), 50°C , 30 min, 90%; b) $(-)\text{-3}$ (1.5 equiv), DDQ (2.0 equiv), *o*-dichlorobenzene, 150°C , 1 h, 61%; c) BBr_3 (8.0 equiv) CH_2Cl_2 , -78°C to RT, 4 h, 86%. DDQ = 2,3-dichloro-5,6-dicyanobenzoquinone, DMSO = dimethyl sulfoxide, IBX = *o*-iodoxybenzoic acid.



Scheme 5. Reagents and conditions: a) H-Cube (Pd/C), H_2 (40 bar), MeOH (0.03 M), 50°C , 0.5 mL min^{-1} , quantitative; b) $\text{LiOH}\cdot\text{H}_2\text{O}$ (1.0 equiv), EtOH/toluene, azeotrope; $\text{Cu}(\text{CH}_3\text{CN})_4\text{PF}_6$ (2.2 equiv), $(-)\text{-sparteine}$ (2.3 equiv), 3 Å molecular sieves, O_2 , THF, -78°C , 16 h; c) benzene, reflux, 12 h, 47% (over 2 steps). H-Cube = continuous-flow hydrogenation reactor, THF = tetrahydrofuran.

displayed robust selectivity with a mean GI_{50} value of $0.21\text{ }\mu\text{M}$. COMPARE analysis of the data^[21] at the total growth inhibition level suggested that $(+)\text{-2}$ might act through interference with tubulin function, as high correlations were seen to the data for seven established tubulin inhibitors.^[10] Examination of this hypothesis by using an *in vitro* tubulin polymerization assay^[22] found this to be the case, with an IC_{50} value of $2.0 \pm 0.1\text{ }\mu\text{M}$, while $(-)\text{-2}$ had no effect at $40\text{ }\mu\text{M}$. $(+)\text{-2}$ was also tested for inhibition of colchicine binding,^[23] where it was found to have moderate activity at $50\text{ }\mu\text{M}$ with $5\text{ }\mu\text{M}$ [^3H]colchicine and $1\text{ }\mu\text{M}$ tubulin. Finally, we confirmed that $(+)\text{-2}$ had an effect on cells consistent with its inhibitory effects on the assembly of tubulin. A cytotoxic concentration of $(+)\text{-2}$ arrested cells in mitosis concordant with inhibition of cell growth (Figure 1) and caused the disassembly of the intracellular microtubule network (Figure 2).^[24]

In conclusion, we have accomplished the total syntheses of $(+)\text{-}$ and $(-)\text{-}$ chamaecypa- none C. The key transformation involved a Diels–Alder cycloaddition between a diarylcyclopentadienone, which was generated *in situ*, and a chiral *ortho*-quinol derived from a retro-Diels–Alder reaction of its dimeric form. Initial biological studies indicate that $(+)\text{-}$ chamaecypa- none C is a potent tumor cell growth inhibitor^[3] that acts primarily through inhibition of tubulin polymerization. Further studies

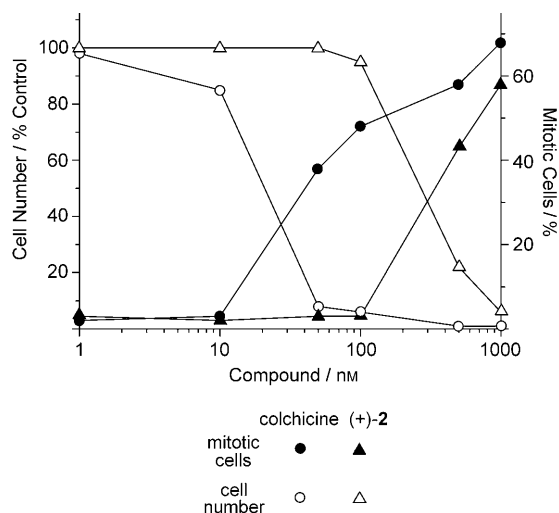


Figure 1. Human Burkitt lymphoma CA46 cells, obtained from the American Type Tissue Collection, were grown in a suspension culture for 24 hours at 37°C in a humidified, 5% CO_2 atmosphere. The medium was RPMI 1640 supplemented with 5% fetal bovine serum. Initially, the culture medium contained $20000\text{ cells mL}^{-1}$. For determination of the cell growth, the increase in cell number was determined with the cells counted in a model Z1 particle counter (Beckman Coulter). For determination of mitotic cells, cells were harvested by centrifugation, briefly swollen in a hypotonic solution, fixed on a glass slide, and stained with Giemsa. The percentage of cells with condensed chromosomes was then determined.

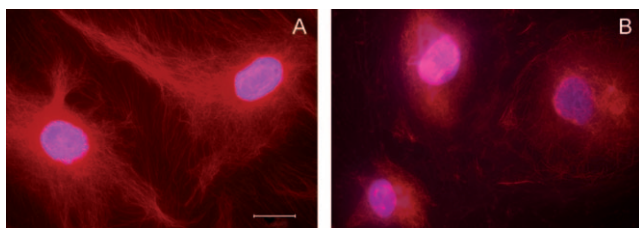


Figure 2. Disruption of the intracellular microtubule network by chamaecypanone C. *Potorus tridactylis* PtK2 kidney epithelial cells were obtained from the American Type Culture Collection and were cultured in minimal essential medium supplemented with 10% fetal bovine serum, 1 mM glutamine, and 1 mM sodium pyruvate. The cells were grown to confluence, disrupted by trypsinization, and seeded at about 35 000 cells into each compartment of a chambered coverglass system (Nunc) with A) no compound or B) 0.5 μM of (+)-chamaecypanone C added to the culture medium. After growth for 16 h at 37°C in a humidified, 5% CO_2 atmosphere, the cells on the coverglass were fixed with acetone at -20°C , washed with phosphate-buffered saline, and stained with a DNA stain and with a monoclonal antibody to β -tubulin conjugated to the fluorescent dye Cy3 (Sigma product C4585: instructions provided by the manufacturer were followed). The coverglass was mounted on a slide with antifade mounting solution and examined in a Nikon Eclipse E800 microscope with a 100-times oil objective and by using appropriate epifluorescence accessories. Images were captured with a Spot digital camera. The scale bar shown in A) represents 20 μm .

on the preparation of (+)-chamaecypanone C analogues by using a retro-DA/DA cascade process, as well as biological evaluation of these compounds, are currently in progress and will be reported in due course.

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- [1] For reviews on biosynthetic Diels–Alder reactions, see: a) E. M. Stocking, R. M. Williams, *Angew. Chem.* **2003**, *115*, 3186–3223; *Angew. Chem. Int. Ed.* **2003**, *42*, 3078–3115; b) H. Oikawa, T. Tokiwano, *Nat. Prod. Rep.* **2004**, *21*, 321–352.
- [2] Y. H. Kuo, C. H. Chen, S. L. Huang, *Chem. Pharm. Bull.* **1998**, *46*, 181–183.
- [3] S. C. Chien, J. Y. Chang, C. C. Kuo, C. C. Hsieh, N. S. Yang, Y. H. Kuo, *Tetrahedron Lett.* **2007**, *48*, 1567–1569.
- [4] R. M. Carman, L. K. Lambert, W. T. Robinson, J. M. A. M. Van Dongen, *Aust. J. Chem.* **1986**, *39*, 1843–1850.
- [5] S. Al-Busafi, J. R. Doncaster, M. G. B. Drew, A. C. Regan, R. C. Whitehead, *J. Chem. Soc. Perkin Trans. 1* **2002**, 476–484.
- [6] S. Dong, J. Zhu, J. A. Porco, Jr., *J. Am. Chem. Soc.* **2008**, *130*, 2738–2739.
- [7] For use of *ortho*-quinol dimers in a retro-DA/DA cascade, see: a) V. K. Singh, P. T. Deota, A. V. Bedekar, *J. Chem. Soc. Perkin Trans. 1* **1992**, 903–912; b) V. Singh, B. Samanta, *Tetrahedron Lett.* **1999**, *40*, 1807–1810; for use of MOB-derived dimers, see: c) C. C. Liao, R. K. Peddinti, *Acc. Chem. Res.* **2002**, *35*, 856–866;
- d) S. K. Chittimalla, H. Y. Shiao, C. C. Liao, *Org. Biomol. Chem.* **2006**, *4*, 2267–2277.
- [8] For reviews on *ortho*-quinol and MOB-derived [4+2] cyclo-dimers, see: a) D. Magdziak, S. J. Meek, T. R. R. Pettus, *Chem. Rev.* **2004**, *104*, 1383–1430; b) S. Quideau, L. Pouysegue, D. Deffieux, *Synlett* **2008**, 467–495.
- [9] B. B. Snider, Z. Chen, *Org. Prep. Proced. Int.* **1999**, *31*, 537–541.
- [10] See the Supporting Information for complete experimental details.
- [11] D. Hafner, K. Goliash, *Angew. Chem.* **1960**, *72*, 781.
- [12] M. Prashad, J. C. Tomesch, J. R. Wareing, H. C. Smith, S. H. Cheon, *Tetrahedron Lett.* **1989**, *30*, 2877–2880.
- [13] S. Datta, A. Odedra, R. S. Liu, *J. Am. Chem. Soc.* **2005**, *127*, 11606–11607.
- [14] J. Szymoniak, J. Besancon, A. Dormond, C. Moise, *J. Org. Chem.* **1990**, *55*, 1429–1432.
- [15] K. C. Nicolaou, Y. L. Zhong, P. S. Baran, *J. Am. Chem. Soc.* **2000**, *122*, 7596–7597.
- [16] a) D. R. Buckle in *Encyclopedia of Reagents for Organic Synthesis*, Vol. 3 (Ed.: L. A. Paquette), Wiley, Chichester, **1995**, pp. 1699; b) D. Walker, J. D. Hiebert, *Chem. Rev.* **1967**, *67*, 153–196.
- [17] For syntheses of cyclopentadienones from cyclopentenones, see: a) P. G. Baraldi, A. Barco, S. Benetti, G. P. Pollini, E. Polo, D. Simoni, *J. Chem. Soc. Chem. Commun.* **1984**, 1049–1050; b) M. Harmata, C. L. Barnes, J. Brackley, G. Bohnert, P. Kirchhoefer, L. Kuerti, P. Rashatasakhon, *J. Org. Chem.* **2001**, *66*, 5232–5236, and references therein.
- [18] For representative syntheses of cyclopentadienones, see: a) P. A. Wender, T. J. Paxton, T. J. Williams, *J. Am. Chem. Soc.* **2006**, *128*, 14814–14815, and references therein; for preparation and studies on 2,4-di-*tert*-butylcyclopentadienone, see: b) E. W. Garbisch, Jr., R. F. Sprecher, *J. Am. Chem. Soc.* **1969**, *91*, 6785–6800; c) R. C. DeSelms, W. R. Schleigh, *Synthesis* **1973**, 614–615; for a total synthesis of the 2,3-substituted cyclopentadienone-derived natural product manzamenone A, see Ref. [5].
- [19] The presumed cyclopentadienone **28** was also successfully trapped with dimethyl acetylenedicarboxylate to afford a tetrasubstituted aromatic product.^[10] For examples of cyclopentadienones as 4π partners in cycloadditions, see: a) T. Ban, Y. Wakita, K. Kanematsu, *J. Am. Chem. Soc.* **1980**, *102*, 5415–5416; b) T. Jikyo, M. Eto, K. Harano, *J. Chem. Soc. Perkin Trans. 1* **1998**, 3463–3470.
- [20] For deantiaromatization in electrocyclic reactions, see: a) M. Harmata, P. Zheng, P. R. Schreiner, A. Navarro-Vázquez, *Angew. Chem.* **2006**, *118*, 2000–2005; *Angew. Chem. Int. Ed.* **2006**, *45*, 1966–1971; for use of reactive cyclopentadienones in cycloaddition, see: b) M. Harmata, M. G. Gomes, *Eur. J. Org. Chem.* **2006**, 2273–2277.
- [21] K. D. Paull, C. M. Lin, L. Malspeis, E. Hamel, *Cancer Res.* **1992**, *52*, 3892–3900.
- [22] E. Hamel, *Cell Biochem. Biophys.* **2003**, *38*, 1–21.
- [23] P. Verdier-Pinard, J.-Y. Lai, H.-D. Yoo, J. Yu, B. Marquez, D. G. Nagle, M. Nambu, J. D. White, J. R. Falck, W. H. Gerwick, B. W. Day, E. Hamel, *Mol. Pharmacol.* **1998**, *53*, 62–72.
- [24] Chang and coworkers (National Health Research Institutes, Taipei, Taiwan) have also independently demonstrated that chamaecypanone C is a microtubule inhibitor, see: C. C. Hsieh, Y. H. Kuo, C. C. Kuo, C. Y. Chang, S. C. Chien, J. Y. Chang, *The 13th Joint Congress of Oncology Societies of Taiwan* (Taipei, Taiwan), May 3–4, **2008** (Abstract A-I-18).